

DETAILED ACTION

This Office action is in response to the communication filed 8-24-10.

Claims 23, 26, 28, 30-38, 40-43, 61-78 are pending in the instant application.

Response to Arguments and Amendments

Withdrawn Rejections

Any rejections not repeated in this Office action are hereby withdrawn.

Maintained Rejections

Claim Rejections - 35 USC § 112

The following is a quotation of the first paragraph of 35 U.S.C. 112:

The specification shall contain a written description of the invention, and of the manner and process of making and using it, in such full, clear, concise, and exact terms as to enable any person skilled in the art to which it pertains, or with which it is most nearly connected, to make and use the same and shall set forth the best mode contemplated by the inventor of carrying out his invention.

Claims 23, 26, 28, 30-38, 40-43, 61-78 are rejected under 35 U.S.C. 112, first paragraph, because the specification, while being enabling for the in vitro increase in splicing efficiency by enhanced inclusion of exon 7 within SMN2 using particularly described 5'GGA and 5'GAA containing antisense oligonucleotides of SEQ ID NOs. 8 and 10, does not reasonably provide enablement for methods of recruiting RNA splicing factors, enhancing exonic incorporation, and recruiting the instantly claimed splicing factors to any target RNA species for treating any condition characterized by defective RNA splicing in an individual, comprising the administration of a representative number

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of species of the broad genus of compounds claimed for the reasons of record set forth in the Office action mailed 2-26-10, and as set forth below.

Applicant's arguments filed 8-24-10 have been fully considered but they are not persuasive. Applicant argues that determining accessible target sites on a target RNA molecule is standard for any method involving annealing of polynucleotides, and even if the optimal mode of delivery has to be determined for individual specific indications before clinical approval is obtained, this would not detract from enablement of the instant claims. In support of the predictability of delivery, Applicant provided the publication by Hua, which involved direct injection of antisense into the CNS of mice in vivo. Applicant also argued that enablement standards have been improperly applied to the instant claims, because they require demonstration that the claimed invention must be useful in a clinical setting. Applicant also argued that The Office action was incorrect in its enablement analysis because the 5'GAA was found to be successful, while the 5'GAA was not.

The claims are broadly drawn to methods of recruiting RNA splicing factors to any target RNA species for treating any condition characterized by defective RNA splicing in an individual, comprising the administration of a nucleic acid molecule comprising a first domain which forms a specific binding pair with any target sequence which is within 100 nucleotides of any defective RNA splicing site on a target RNA, and which nucleic acid molecule further comprises a second specific binding pair with the specifically claimed RNA splicing factors and which second domain is non-complementary to the second target RNA species.

Contrary to Applicant's assertions, the direct administration of an antisense into the CNS of a mouse does not provide enablement for the ability to treat any condition characterized by defective RNA splicing in an individual comprising administration, by any means, of the expansive genus of compounds claimed. Applicant is correct that the specification teaches an increase in the proportion of exon 7 inclusion in the SMN2 mRNA using 5'GGA and GAA oligonucleotides, but the effect of the 5' GAA was largely accounted for by a decrease in the level of skipped mRNA. The specification also teaches the characterization of a series of oligonucleotides with tails for recruiting hTRA2 β and SF2/ASF, whereby a specific increase in the proportion of exon 7 inclusion in SMN2 mRNA was observed in vitro when using 5'GAA but not with 5'GGA. See, e.g., page 65 of the instant disclosure, teaching that hTra2-B1 is thought to be rendered inaccessible to 5'GGA due to possible secondary structural constraints. See also page 66, showing that the addition of hTra2B to the nuclear extract stimulates inclusion in SMN1, but has relatively little effect on SMN2 in the presence of either 5'GGA or 5'GAA, suggesting that other factors limit improvements in corrective efficiency that appear unpredictable. These results illustrate the unpredictability in the field of splicing factor recruitment in designing therapeutic candidates for treating conditions in an individual which is characterized by defective or undesirable RNA splicing.

Contrary to Applicant's assertions, the references cited in support of the enablement rejection accurately reflect the unpredictability in the art of gene therapy that remains today, and the success of a particular oligonucleotide to provide in vivo effects after direct injection into the CNS is not an indicator for predictable success for a different

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effector molecule (e.g. a splicing factor) provide correct splicing in an organism exhibiting defective or undesirable RNA splicing.

One is not necessarily able to extrapolate the success in one clinical situation to another situation, especially where different effector molecules are used to target different genes of interest, or are involved in different biochemical mechanisms.

The unpredictability of the field has been illustrated by the instant disclosure (see above) and by others in the field, as cited in the prior Office action. Meyer et al, Human Molec. Genetics, Vol. 18, pages 546-555, 2009, at page 546 of discloses the unpredictability of this field, where Exon 7 of the SMN2 gene is one of the best studied exons of the human genome, yet many hurdles still exist in achieving clinical efficacy: ...”the oligonucleotides did not reach the spinal cord, and hence no therapeutic benefit could be demonstrated...”

Marquis et al, Molecular Therapy, Vol. 15, No. 8, pages 1479-1486, 2007, at page 1483, also addresses the unpredictability of this field:

In all four target locations, the addition of the ESE led to a certain let-up in this exclusion effect. This strongly suggests that the enhancer/U7 combination can be used for improving the recognition of many exons in other genes too. **However, our results also demonstrate the necessity of carefully selecting and optimizing the target locations as well as the ESE sequences.** Only in two instances, i.e., the exonic locations A and B, did the stimulatory effect of the ESE overcome the inhibition caused by the control constructs and lead to a real improvement of exon 7 inclusion. Moreover, targeting of the bifunctional U7 snRNA to position B was more efficient than to position A. The differences in the expression levels of the U7 snRNAs can only partly explain these different stimulatory efficiencies...

(citations omitted, emphasis added).

See also Marquis et al at page 1479, last full paragraph, pointing out the ongoing delivery obstacles that must be overcome in gene therapy: "However, it is unclear whether such oligonucleotides can be delivered to motoneurons, a treatment which would have to be repeated frequently."

The specification fails to teach successful *in vitro* and *in vivo* splicing corrections using a representative number of species of the broadly claimed genus, which encompasses a first domain which forms a specific binding pair with any target sequence which is within 100 nucleotides of any defective RNA splicing site on a target RNA, and which nucleic acid molecule further comprises a second specific binding pair with the specifically claimed genus of RNA splicing factors, which is optionally a UsnRNP. The specification also fails to teach a representative number of species of nucleic acids providing therapeutic effects for any disease or condition characterized by defective or undesirable RNA splicing in an individual. The examples provided at the time of filing are not representative or correlative of the expansive genus of compounds claimed, and would require undue experimentation beyond that existing in the art, or in the instant disclosure, to practice the full scope claimed.

The quantity of experimentation required to practice the invention as claimed would require the *de novo* determination of a representative number of compounds claimed, whereby exonic incorporation has been enhanced, and recruitment has occurred for the claimed genus of splicing factors to any target RNA species for treating any condition characterized by defective RNA splicing in an individual. Other experimentation required to practice the invention claimed includes determining accessible target sites,

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modes of delivery and formulations to target appropriate cells and /or tissues in an organism, whereby the compound or compounds are effectively delivered in adequate quantities to the target cells.

Since the specification and the art fail to provide sufficient guidance for the methods using the therapeutic compositions claimed, and since determination of these factors is highly unpredictable, it would require undue experimentation to practice the invention over the broad scope claimed. Thus, the full scope is not enabled.

For these reasons, the instant rejection is maintained.

New Rejections

Applicant's arguments with respect to the previous 103 rejection have been considered but are moot in view of the new ground(s) of rejection set forth below.

Claim Rejections - 35 USC § 103

The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:

(a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negated by the manner in which the invention was made.

Claims 23, 26, 28, 30-38, 40-43, 61-69, 71-78 are rejected under 35 U.S.C. 103(a) as being unpatentable over by Carlo et al (Molec. & Cell. Biol., Vol. 20, No. 11, pages 3988-3995, 2000) and Mitchell et al (US 2004/0126774), the combination in view of Hofmann et al (Proc Natl. Acad. Sci., Vol 97, No. 17, pages 9618-9623, 2000, provided by Applicant)), Lim et al (J. Biol. Chem., Vol. 276, No. 48, pages 45,476-

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45,483, 2001, see IDS document No. 61, filed 12-30-05), and Lorson et al (Proc. Natl. Acad. Sci., Vol. 96, pages 6307-6311, 1999, see IDS document No. 64, filed 12-30-05) insofar as the claims are drawn to methods of recruiting RNA splicing factors to an RNA species and enhancing exonic incorporation in vitro comprising providing a nucleic acid molecule which optionally comprises stabilizing modifications including 2-O-methyl modified sugars or phosphorothioate internucleotide linkages, and having a first nucleotide sequence capable of forming a first specific binding pair with a target sequence via complementary base pairing, and a second nucleotide sequence capable of forming a second specific binding pair with an RNA splicing factor selected from SR, SR-related, hnRNP and U1 snRNP proteins, which second nucleotide sequence is not complementary to the target RNA species, wherein the two binding pairs form to recruit the RNA splicing factor to an RNA splicing site at the RNA splicing site of the target RNA, which second sequence is optionally CAGGUAAGU, whereby cis-splicing is optimized for treatment of a condition characterized by defective RNA splicing in an individual, and which target RNA species is optionally SM1 or SM2, and which splicing factor is SF2/ASF.

Carlo et al (Molec. & Cell. Biol., Vol. 20, No. 11, pages 3988-3995, 2000) (See Reference No. 18 in the IDS filed 12-30-05) teach methods of bridging exons for enhancing cis-splicing by exploiting the appropriate binding elements involved in splicing machinery components, which components are required for bringing the splicing components to the proximal distances required for enhanced exon inclusion in

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the splicing process. Carlo also teaches the role of U1snRNP's in exon recognition (see entire article, esp. the text on p. 3988-9, figure 8, text on p. 3994-5).

Mitchell et al (US 2004/0126774) teach methods of recruiting RNA splicing factors, enhancing exonic incorporation, and recruiting a splicing factor to a target RNA species, comprising the administration of a nucleic acid molecule comprising a first domain which forms a specific binding pair with a target sequence which is within 100 nucleotides of a defective RNA splicing site on a target RNA, and which nucleic acid molecule further comprises a second specific binding pair with an RNA splicing factor which is optionally a UsnRNP, and which second domain is optionally complementary or non-complementary to the second target RNA species, and optionally comprises CAGGUAAGU which represents the well known 5' splice site consensus sequence involved in cis and trans-splicing (see the abstract; pages 1-6, esp. ¶¶ 0008-0009, 0013, 0015, 0016, 0019-0020, 0037, 0039, 0040, 0042-0044, 0047-0051, 0062, 0065, claims 1-11, 16-36).

Mitchell also teaches the well known similarities between cis-splicing and trans-splicing at ¶0006:

The mechanism of splice leader trans-splicing which is nearly identical to that of conventional cis-splicing, proceeds via two phosphoryl transfer reaction. The first causes the formation of a 2'-5'phosphodiester bond producing a 'Y' shaped branched intermediate, equivalent to the lariat intermediate in cis-splicing. The second reaction, exon ligation, proceeds as in conventional cis-splicing. In addition, sequences at the 3' splice site and some of the snRNPs which catalyze the trans-splicing reaction, closely resemble their counterparts involved in cis-splicing.

The primary references do not teach the role of improper splicing of the SMN gene in spinal muscular atrophy, nor do they teach the incorporation of 2'-O-methyl or phosphorothioate internucleotide modifications into oligonucleotides.

Hofmann et al (Proc Natl. Acad. Sci., Vol 97, No. 17, pages 9618-9623, 2000) teach promotion of the inclusion of SMN2 exon 7 in stimulation of full length SMN2 expression in vitro upon transfection of target cells in vitro using commercially available liposomes, and subsequent expression of recombinant Htra2-Beta1, which is an SR-like splicing factor which was found to bind an AG-rich exonic splicing enhancer in SMN exon 7 (see entire document, esp. the text on p. 9618; right hand col. on p. 9619; text on p. 9622).

Lorson et al (Proc. Natl. Acad. Sci., Vol. 96, pages 6307-6311, 1999, see IDS document No. 64, filed 12-30-05) teach the single nucleotide mutation in the splice site of the SMN gene which regulates splicing and teaches a direct relationship between this single nucleotide mutation, the presence of disease and exon 7 skipping in SMN (see esp. the abstract and introduction on p. 6307, discussion on pages 6310-6311).

Lim et al (J. Biol. Chem., Vol. 276, No. 48, pages 45,476-45,483, 2001, see IDS document No. 61, filed 12-30-05) teach the single nucleotide mutation in the splice site of the SMN gene which leads to the disruption of an exonic splicing enhancer and reduces the strength of the 3'-splice site of exon 7, reduces intron 6 removal, and increases the efficiency of competing exon 7 skipping pathway, which results in progression of spinal muscular atrophy. Lim also teaches enhanced stability, target binding and cellular uptake of oligonucleotides comprising 2'-O-methyl and phosphorothioate internucleotide modifications (see the abstract, introduction on pages

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45,476-45,477; methods on page 45,478; fig. 2 on p. 45,479; discussion on pages 45,480, and 45,482-45,483).

It would have been obvious to recruit RNA splicing factors to a target RNA species as instantly claimed to enhance cis-splicing because the general phenomenon of enhancing splicing by bringing the necessary splicing components within close proximity to each other, and to a splicing site were well known in the art as taught previously by Carlo, where binding partners were exploited to bring the necessary factors together to alter or enhance exon splicing. It would have been obvious to utilize complementary sequences of known splicing factors as a means of localizing required factors because it was well known in the art that nucleic acids bind to their complementary sequences. It was also well known in the art that U1 snRNP bound to the 5' splicing site or pre-mRNA, and is an essential factor for cis-splicing. It was also well known that the motif CAGGUAAGU represents the well known 5' splice site consensus sequence and is exploitable for use as a second binding pair for localizing the necessary components for cis-splicing.

One would have been motivated to apply the teachings of Carlo and Mitchell to correct the splicing defect involved in spinal muscular atrophy by enhancing the efficiency of the endogenous exon 7, and reduce intron 6 removal in survival motor neuron (SMN1) alleles because it was well known that spinal muscular atrophy is caused by exon 7 skipping, leading to the loss of functional SMN1 alleles and leading to expression of a non-functional gene product due to differential SMN2 splicing caused by disruption of an exonic splicing enhancer. One therefore would have been motivated to

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design a nucleic acid molecule comprising a nucleic acid sequence that is a complementary sequence to well known crucial splicing component that is near to the splicing site, and a second binding partner to localize the splicing components U1, SF2/ASF to the splice site in order to optimize steric proximity and the accessibility of these necessary splicing components for enhancing cis-splicing. It would also have been obvious to design nucleic acid molecules comprising a first domain which forms a specific binding pair with a target sequence of SMN that is within 100 nucleotides of the defective RNA splicing site taught previously by Lorson and Lim, and which nucleic acid molecules further comprise a second specific binding pair with an RNA splicing factor appropriate for recruiting the relevant RNA splicing factors, for reducing exon 7 skipping or optionally for enhancing exonic incorporation, and for recruiting splicing factors to the target SMN RNA for treating spinal muscular atrophy (SMA) because the mutation linking this disease with incorrect or aberrant splicing was well known in the art at the time the instant invention was made. Furthermore, it would have required routine experimentation to design nucleic acid molecules targeting the mutation site previously taught in the art for SMN and relying on the methods taught previously by Hofmann, Mitchell and Carlo.

One of ordinary skill in the art would have had a reasonable expectation of success for correcting the exon skipping phenomenon in vitro associated with SMA because the single point mutation causing this exon skipping was well known in the art, and the design of nucleic acid molecules for targeting and correcting this mutation were also well known in the art, relying on the teachings of Hofmann, Mitchell and Carlo,

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whereby administration of recombinant Htra2-Beta 1, or administration of splicing molecules designed to interact with the well known target precursor messenger RNA molecule of SMN would reasonably be expected to lead to trans-splicing to correct the mutated splice site in appropriate target cells in vitro.

One of ordinary skill in the art would have been motivated to correct this genetic defect because the relationship between exon skipping and SMA disease severity had been well documented the prior art, and the means of generating pre-trans-splicing molecules to trans-splice the target pre-mRNA were also well known in the art to provide gene therapy approaches for correcting splicing disorders for known mutations.

One of ordinary skill in the art would also have been motivated to incorporate 2'-O-methyl and/or phosphorothioate internucleotide modifications into nucleic acid molecules for target cell delivery and uptake because these modifications were well known in the art to enhance nucleic acid stability from nuclease degradations, as well as enhancing target cell uptake and target binding, as taught previously by many in the field, including Lim et al.

For these reasons, the instant invention would have been obvious to one of ordinary skill at the time the instant invention was made.

Allowable Subject Matter

Seq ID No. 16 (recited in claim 70) appears free of the prior art searched and of record.

Conclusion

Certain papers related to this application may be submitted to Art Unit 1635 by facsimile transmission. The faxing of such papers must conform with the notices published in the Official Gazette, 1156 OG 61 (November 16, 1993) and 1157 OG 94 (December 28, 1993) (see 37 C.F.R. ' 1.6(d)). The official fax telephone number for the Group is **571-273-8300**. NOTE: If Applicant *does* submit a paper by fax, the original signed copy should be retained by applicant or applicant's representative. NO DUPLICATE COPIES SHOULD BE SUBMITTED so as to avoid the processing of duplicate papers in the Office.

Any inquiry concerning this communication or earlier communications from the examiner should be directed to **Jane Zara** whose telephone number is **(571) 272-0765**. The examiner's office hours are generally Monday-Friday, 10:30am - 7pm. If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Heather Calamita, can be reached on (571) 272-2876. Any inquiry of a general nature or relating to the status of this application should be directed to the Group receptionist whose telephone number is (703) 308-0196.

Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see <http://pair-direct.uspto.gov>. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free).

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